

# Original Article



## Toxicity of nanoZnO in *Daphnia magna* fed with ZnO containing *Chlorella vulgaris* and *Scenedesmus dimorphus* algae

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### Abstract

Nano ZnO is currently used in the rubber, electronics, electrical appliances, enamel, cosmetics and medical industry. Whereas most studies have used the ecological toxicity of nanoparticles , the toxic effects of nanoparticles in diet is not extensively explored. Because the algae are at the base of the food chain, any change in their density, biomass and population, would affect the food chain in aquatic ecosystems. *Daphnia magna* is widely used in environmental pollution control measures, especially in nanoparticle toxicity tests for its short reproduction time and high sensitivity. In this study, *Chlorella vulgaris* and *Scenedesmus dimorphus* algae were contaminated with nano ZnO. Five treatment concentrations and three replications at any concentration were compared between test and control groups and the results were analyzed by Probit method. LC<sub>50</sub> of *Chlorella vulgaris* and *Scenedesmus dimorphus* algae fed by nano ZnO containing *Daphnia magna* was obtained at 1.78 and 2.59 mg/l levels after 96 hours respectively. Moreover the NOEC (No Observed Effect concentration) and LOEC (lowest observed effect concentration) were calculated as 0.17 and 0.25 mg/l for *Scenedesmus dimorphus* algae and *Chlorella* respectively. Our results showed that *Daphnia magna* is more sensitive to a diet comprising nano ZnO fed *Scenedesmus dimorphus* algae as compared to *Scenedesmus dimorphus* environmentally contaminated with nano ZnO.

**Keywords:** Nano ZnO, *Scenedesmus Dimorphus*, *Chlorella Vulgaris*, *Daphnia Magna*, NOEC, LOEC

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## Introduction

Nanotechnology is a fast growing science and nano products developed on a daily basis are continuing their impact on human lifestyle. Limited studies have dealt with the potential dangerous aspects of nanoproducts on ecosystem. One major reason for this scarcity is difficulty of identifying the effects of nanoparticles using currently available test methods. Nanoparticles can enter environment and transmit through air, soil and underground water, potentially affecting the health of organisms living in these environments. In addition, nanoparticles can enter the food chain by micro-organisms such as bacteria. Presence of nanoparticles in living organisms may have a potentially destructive effects leading to irreversible damage to them. While producing and irregular use of these nano materials can ultimately lead to exposure of ecosystems to these substances there is no comprehensive information about the rate of this exposure (Monica et al., 2009).

Among identified features of nanoparticles, mobility via water, absorption by plants, presence in plant tissue and soil, and suspension in the air can be mentioned (Ferry et al., 2009). Because of small sizes, nanoparticles can pass many obstacles and filters. Use of nanoparticles in commercial scale has the potential negative consequences (Nanotechnology., 2006). These particles can easily be transmitted through the wind and because of having large and active surface area, they can stick to heavy metals, chemical and organic materials there by causing soil and surface water pollution. It is well documented that toxic effects of nanoparticles can dramatically increases by reduction of their size and increasing their concentration (Oberdorster et al., 1996. Yang et al., 2005, Lee et al., 2008). Nanoparticles are usually not soluble in aqueous environments; rather they mostly appear in suspended form. It is important to

note that the mobility of nanoparticles in aquatic environments results in the fact that a higher number of marine organisms including algae and fish are exposed to the potential irreparable damages associated with their toxic activity (Corredor et al., 2009).

Among the most widely used nanoparticles, nano ZnO (n-ZnO), has been widely used in rubber, electronics and electrical industries, and in developing cosmetics products, and medical treatments moreover. N-ZnO can lead to generation of free radicals in skin cells, damaging DNA, and altering the structure of proteins, which can potentially lead to promotion of cancer disease (Environmental Protection Agency Nanotechnology, 2007)

There is a limited ecological toxicity dealing with the toxicity of nanoparticles on phytoplankton. This phenomenon showed that surface structure and matrix of cell wall can function as a surface for growth of nanoparticle (Hundrinke., 2006). In addition, some other investigators have identified some forms of stable nano crystals in marine phytoplankton in contact with cadmium. Similar studies dealing with bacteria are very limited. However, antibacterial effects of dioxid titanium and silver nanoparticles are well documented (Duran et al., 2007). It can be therefore expected that these substances have a potential toxicity to other bacteria as well. Oberdorster et al. (2006) have investigated the relationship between the size of silica nanoparticles *Chlorella kessleri* algae and their toxic potentials. The study found an inverse relationship between size and toxicity of silica nanoparticles in this species. (Fujiwara et al., 2008)

While numerous studies has been carried out on the toxic effects of n-ZnO on living animals, studies regarding transmission of this substance from a surface of food chain to another is limited. Given this limitation, the present study was focused on the impact of n-ZnO on algae *Scenedesmus dimorphus* and *Chlorella vulgaris* as two producers of *Daphnia magna*.

## Materials and Methods

In this study, neonates with less than 24 hours old were used according to the recommendations of OECD<sup>1</sup> (OECD Guideline for testing of Chemicals. 2002, )

In order to bearing 24-hour-old neonates, female *Daphnia* containing eggs were isolated. The female *Daphnia* were then fed with yeast extracts and *Scenedesmus dimorphus* algae. When the eggs hatched, neonates of less than 24 hours old were used in toxicity testing.

During the test period, pH and temperature was controlled at=7-8 and 22±2 °C, respectively. After finding the appropriate ranges, concentration treatments were calculated in logarithmic scale.

*Scenedesmus dimorphus*, *Chlorella vulgaris* algae were exposed to 0, 5, 10, 50 and 100 mg/l of n-ZnO for three days. Afterwards,

in separate treatments these algae were used in nutrition for *Daphnia* the loosing rate of *Daphnia* fed with n-ZnO containing algae, in comparison with controls, was recorded in every 24 hours. Then LC10, LC50, and LC90 values for *Daphnia* feeding with n-ZnO containing algae were calculated using Probit analysis. A significant level of 0.05 was considered in all tests.

## Results

After 96 hours, the daily surveys, with were recorded and analyzed using probit method. R2 values obtained from regression analysis, are shown in Table 1. Lethal doses of each alga contaminated with n-ZnO was calculated at different times.

**Table 1: Regression Curve of nano ZnO based on the concentration algae for *Daphnia***

	<b>h(time)</b>	<b>24</b>	<b>48</b>	<b>72</b>	<b>96</b>
Sc	Line Equation	y=2.056x+1.921	y=0.883x+4.861	y=2.532x+4.362	y=2.532x+4.362
	R <sup>2</sup>	0.927	0.989	0.971	0.971
Ch	Line Equation	y=3.682x+0.179	y=4.356x+1.387	y= 3.125x+3.369	y=2.898x+3.802
	R <sup>2</sup>	0.913	0.998	0.986	0.998

## Toxicity of Chlorellan-ZnO containing algae on *Daphnia magna*

During test period, mortality rate of *Daphnia* due to being feed with n-ZnO containing *Chlorella* valgae was recorded (Figure1). The results of acute toxicity of n-ZnO containing *chlorella* algae on *Daphnia magna* being fed ,

was calculated based on lethal concentration (LC) (Table 2). As seen, after 96-h, a LC10of 0.93, LC50of 2.59 and LC90of 7.17 mg/liter were recorded for n-ZnO algae. Figure 2 comoares the lethal doses on different days. In this test, a NOEC of 0.25 mg/l and a MAC of 0.25 mg/l were calculated.

1. Organization for Economic Cooperation and Development

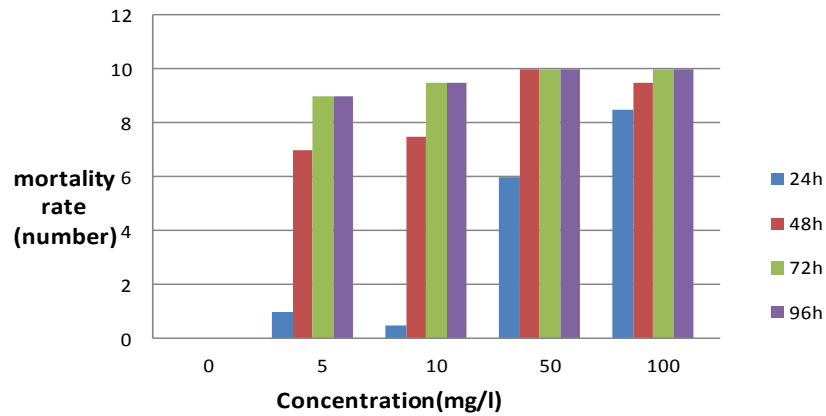


Figure 1: Mortality rate of Daphnia fed algae chlorella contains nano ZnO

Table 2: Lethal dose of chlorella algae containing nano ZnO on Daphnia magna

Time(h)	24	48	72	96
LC <sub>10</sub>	7.33	0.05	0.55	0.55
LC <sub>50</sub>	34.08	1.43	1.78	1.78
LC <sub>90</sub>	158.37	40.62	5.72	5.72

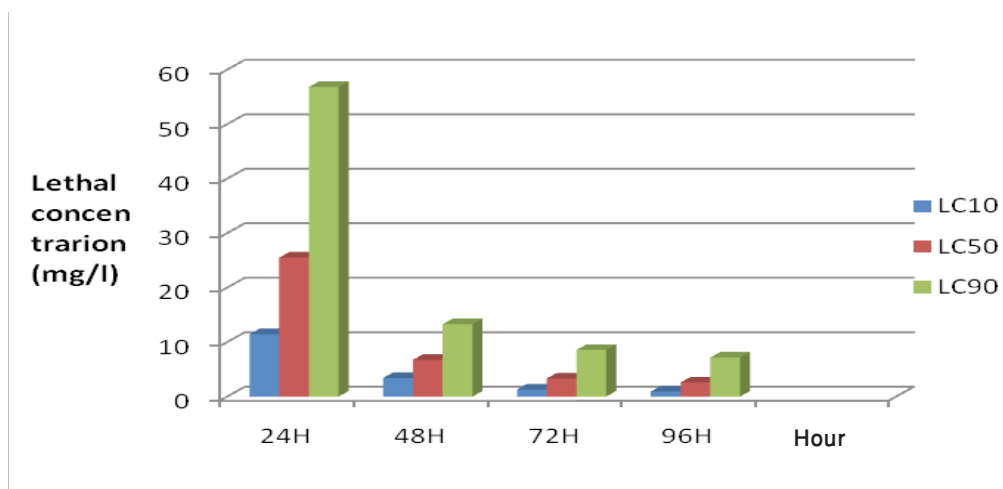


Figure 2: Comparison of lethal doses Chlorella algae containing nano ZnO on Daphnia during 96 hours

**Toxicity of algae *Scenedesmus dimorphus* containing n-ZnO**

Figure 3 shows the mortality rate of *Daphnia magna* due to being fed by n-ZnO containing algae *Scenedesmus dimorphus*. As concentration of n-ZnO and exposure time increased, the mortality rate of *Daphnia magna* increased as well. Toxicity of n-ZnO *Scenedesmus dimorphus*

was calculated in terms of lethal doses per day. LC50 of these n-ZnO containing algae was calculated after 96h to be 1.78 mg/l. As seen in Table 3 after 72 hours, the sensitivity of *Daphnia magna* to n-ZnO containing algae was remained unchanged. In addition, in this test, a NOEC of 0.17 mg/l and a MAC of 0.17 mg/l was calculated.

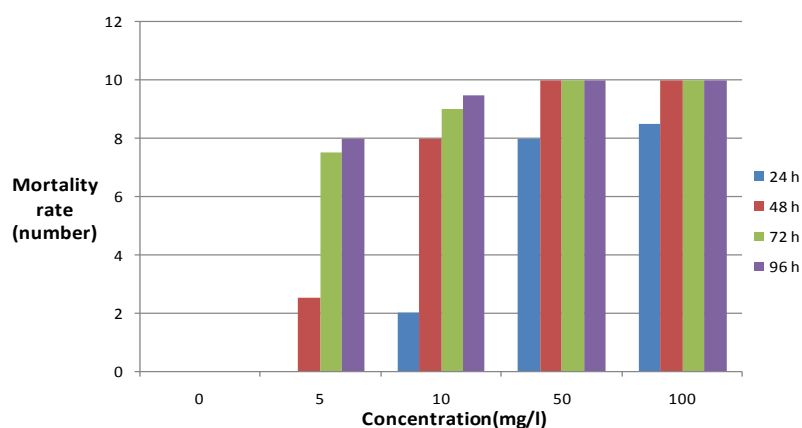


Figure 3. *Daphnia magna* mortality due to be fed by *Scenedesmus dimorphus* containing nano ZnO

Table 3: Lethal dose of algae *Scenedesmus dimorphus* containing nano ZnO on *Daphnia magna*

Time(h) LC(mg/l)	24	48	72	96
LC <sub>10</sub>	7.33	0.05	0.55	0.55
LC <sub>50</sub>	34.08	1.43	1.78	1.78
LC <sub>90</sub>	158.37	40.62	5.72	5.72

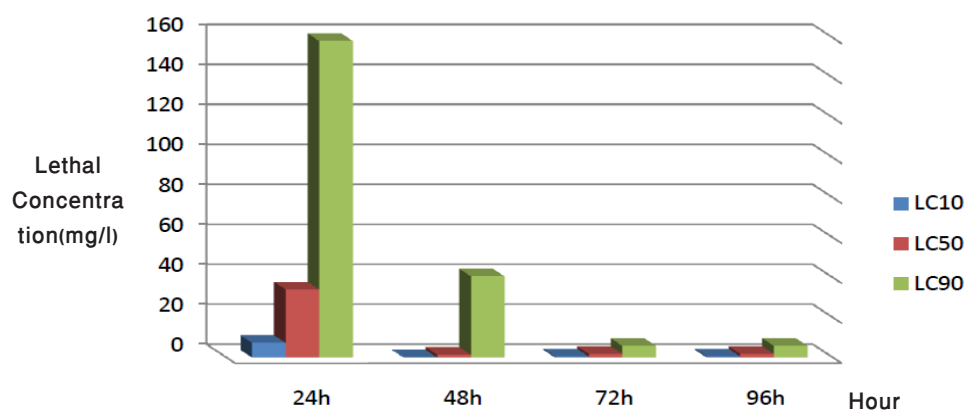


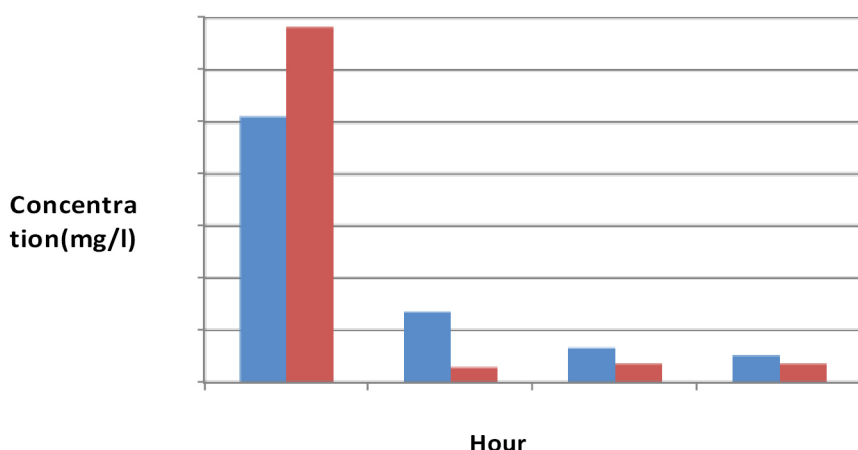
Figure 4: Comparison of lethal concentrations of algae *Scenedesmus dimorphus* containing nano ZnO

## Discussion

During feeding experiments of *Daphnia* with *Scenedesmus dimorphus*, *Chlorella vulgaris* containing ZnO (algae were exposed to n-ZnO after 72h) at 96 h LC50 silica nanoparticles was calculated to be 2.5 mg/l for algae *Chlorella* and 1.78 mg/l for *Scenedesmus dimorphus*. On the basis of these data, it could be concluded that n-ZnO content of *Scenedesmus dimorphus* alga was higher than that of *Chlorella*. In addition, *Scenedesmus dimorphus* alga showed higher sensitivity as compared with *Daphnia magna*. Figure 3 shows that, after 48 h, the LC10, LC50, and LC90

increased with a high rate. *Daphnia* death rates at different concentrations and times is shown in Figure 4 and represents a higher death rate for *Daphnia* when compared with diagram in Figure 3.

In Figure 4, after 96 h in concentration of 5 mg/l, which was the lowest test dose, a 100% mortality rate was observed. However, the toxicity effect of *Chlorella* algae after 96 h in concentration of 10 mg/l, was relatively lower. Figure 5 shows LC50 of n-ZnO containing *Scenedesmus dimorphus* algae and *Chlorella* on *Daphnia* after 96 hours.



**Figure 5: Compared toxicity of algae containing nano ZnO *Daphnia magna*.**

As seen, *Chlorella* algae, shows a lower LC50 *Daphnia* than *Scenedesmus dimorphus* algae containing n-ZnO after 24h. This observation implies that, *Chlorella* algae is more toxic as compared with *Scenedesmus dimorphus* algae; however as time passed, the toxicity of *Scenedesmus dimorphus* algae containing n-ZnO in *Daphnia magna* is higher. This indicates a higher effectiveness of *Daphnia magna* as preferred food compared with *Scenedesmus dimorphus* algae. Limited studies have been conducted about transmis-

sion of the toxicity of nanoparticles through diet. Federici et al. showed that toxicity of nano TiO<sub>2</sub> through diet in trout can result in intestinal damage. Moreover some biochemical evidences have indicated the presence of oxidative stress in intestinal epithelial cells (Federici et al., 2007). It seems that the physicochemical behavior of nanoparticles affects their absorption potentials to many surfaces, including water, algae, biological membranes, soil and the outer surface of living creatures.

## References

- Corredor E, Testillano PS, Coronado M J, Gonzalez-Melendi P, Fernandez-Pacheco R, Marquina C et al. Nanoparticle penetration and transport in living pumpkin plants: in situ subcellular identification. *BMC Plant Biology*. 2009; 9:1-11.
- Duran N, Marcato PD, Desouza GIH, Alves OL, Esposito E. Antibacterial effect of silver nanoparticles produced by fungal process on textile fabrics and their effluent treatment. *J Biomed Nanotech*. 2007; 3:203-208.
- Federici G, Shaw BJ, Handy RD. Toxicity of titanium dioxide nanoparticles to rainbow trout: Gill injury, oxidative stress, and other physiological effects. *Aquat Toxicol* 2007; 84:415-430.
- Ferry JL, Craig P, Hexel C, Sisco P, Frey R, Pennington PL et al. Transfer of goldnanoparticles from the water column to the estuarine food web. *Nature Nanotechnol*. 2009; 4:441-444.
- Fujiwara K, Suematsu H, Kiyomiya E, Aoki M, Sato M, Moritoki N. Size-dependent toxicity of silica nano-particles to *Chlorella kessleri*. *J Environ Sci Health A Tox Hazard Subst Environ Eng*. 2008; 43(10)1167-1173.
- Lee WM, An YJ, Yoon H, and Kweon HS. Toxicity and bioavailability of copper nanoparticles to the terrestrial plants mung bean (*Phaseolus radiatus*) and wheat (*Triticum aestivum*): plant agar test for water in soluble nanoparticles. *Environ Toxicol and Chem*. 2008; 27:1915-1921.
- Monica RC and Cremonini R. Nanoparticles and higher plants. *Caryologia*. 2009; 2(62) 161-165.
- Nanotechnology; Opportunities and risks for humans and the environment, Umwelt Bundes Amt (UBA) background paper. 2006; Germany.
- Nanotechnology; Health and environmental risks of nanoparticles, BAUA, BFR, UBA. 2006; Germany.
- Oberdorster G. Effects of Ultrafine Particles in the Lung and Potential Relevance to Environmental Particles. In Marijnissen, J.M.C., Gradon, L. (Eds), *Aerosol Inhalation*. Kluwer Academic, Dordrecht. 1996; 165-173.
- Oberdorster E, Zhu SQ, Blickley TM, Clellan-Green P, Haasch ML. Ecotoxicology of carbon-based engineered nanoparticles: effects of fullerene on aquatic organisms. *Carbon* 2006; 44:1112-1120
- Test No. 211: *Daphnia magna* Reproduction Test ,OECD Guideline , 02 Oct 2012.
- U.S. Environmental Protection Agency Nanotechnology White Paper Science Policy Council U.S. Environmental Protection Agency Washington, EPA 100/B-07/001. 2007.
- Van Hoecke K, De Schampheleere K, Van der Meeren P, Lucas S, Janssen C. Ecotoxicity of silica nanoparticles to the green alga *Pseudokirchneriella subcapitata*: importance of surface area. *Environ Toxicol and Chem*. 2009; 27(9)1948-1957.
- Wardok A. Nanotechnology and Regulations, A case study using the Toxic Substance Control Act (TSCA), a discussion paper, foresight and governance project. 2008
- Yang L and Watts DJ, particles surface characteristics may play an important role in phytotoxicity of alumina nanoparticles. *Toxicology Letters*. 2005; 158:122-132.